

Refine Search

Search Results -

Term	Documents
TWO-STAGE	69450
TWO-STAGES	618
MULTIPLEX	115558
MULTIPLEXES	21280
PCR	87660
PCRS	2628
AMPLIFICATION	255073
AMPLIFICATIONS	9853
(TWO-STAGE AND ((MULTIPLEX ADJ PCR) OR (MULTIPLEX ADJ AMPLIFICATION))).PGPB,USPT,USOC,EPAB,JPAB,DWPI.	32
((TWO-STAGE AND (MULTIPLEX PCR OR MULTIPLEX AMPLIFICATION))).PGPB,USPT,USOC,EPAB,JPAB,DWPI.	32

Database:

US Pre-Grant Publication Full-Text Database
 US Patents Full-Text Database
 US OCR Full-Text Database
 EPO Abstracts Database
 JPO Abstracts Database
 Derwent World Patents Index
 IBM Technical Disclosure Bulletins

Search:

L64

Refine Search

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Search History

DATE: Tuesday, February 01, 2005 [Printable Copy](#) [Create Case](#)

Set
Name Query
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 side

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Count

Set
Name
 result
 set

DB=PGPB,USPT,USOC,EPAB,DWPI; PLUR=YES; OP=ADJ

L1 10/090326

1 L1

<u>L36</u>	L35 and (vessel or reaction vessel)	36	<u>L36</u>
<u>L37</u>	L36 and automat\$	25	<u>L37</u>
<i>DB=PGPB,USPT,USOC,EPAB,DWPI; PLUR=YES; OP=ADJ</i>			
<u>L38</u>	intraoperative PCR	2	<u>L38</u>
<u>L39</u>	(operation or surgery) same (PCR or polymerase chain reaction)	4124	<u>L39</u>
<u>L40</u>	L39 same threshold	42	<u>L40</u>
<u>L41</u>	L40 same indicator transcript	1	<u>L41</u>
<u>L42</u>	((operation or surgery)same patient) same (PCR or polymerase chain reaction)	294	<u>L42</u>
<u>L43</u>	L1 and indicator transcript	1	<u>L43</u>
<u>L44</u>	L43 and CEA transcript	0	<u>L44</u>
<u>L45</u>	L1 and CEA transcript	0	<u>L45</u>
<u>L46</u>	L1 and CEA	1	<u>L46</u>
<u>L47</u>	carcinoembryonic antigen	3761	<u>L47</u>
<u>L48</u>	L47 and L42	33	<u>L48</u>
<u>L49</u>	L48 and (during near\$2 (surgery or operation))	0	<u>L49</u>
<u>L50</u>	L48 and (during same(surgery or operation))	19	<u>L50</u>
<u>L51</u>	L42 and (during same(surgery or operation))	119	<u>L51</u>
<u>L52</u>	((PCR or polymerase chain reaction)same (during same(surgery or operation)))	1024	<u>L52</u>
<u>L53</u>	L52 same L47	1	<u>L53</u>
<u>L54</u>	L52 same tumor antigen	0	<u>L54</u>
<u>L55</u>	L52 and L47	3	<u>L55</u>
<u>L56</u>	L52 and transcript	74	<u>L56</u>
<u>L57</u>	L52 same transcript	9	<u>L57</u>
<u>L58</u>	L52 same expression	29	<u>L58</u>
<u>L59</u>	L52 same threshold	6	<u>L59</u>
<u>L60</u>	L47 and L7	31	<u>L60</u>
<i>DB=USPT,DWPI; PLUR=YES; OP=ADJ</i>			
<u>L61</u>	5837442.pn or 5882856.pn. or 5965710.pn. or 6057105.pn. or 6168948.pn.	8	<u>L61</u>
<i>DB=PGPB,USPT,USOC,EPAB,JPAB,DWPI; PLUR=YES; OP=ADJ</i>			
<u>L62</u>	(two-stage near (PCR reaction or amplification reaction))	10	<u>L62</u>
<u>L63</u>	(two-stage near (multiplex PCR or multiplex amplification))	0	<u>L63</u>
<u>L64</u>	(two-stage and (multiplex PCR or multiplex amplification))	32	<u>L64</u>

END OF SEARCH HISTORY

<u>L2</u>	multiplex PCR	1216	<u>L2</u>
<u>L3</u>	L1 and first amplification and second amplification	1	<u>L3</u>
<u>L4</u>	L1 and ((first same second) near (amplif\$ or cycle\$1))	1	<u>L4</u>
<u>L5</u>	L1 and ((different or multiple or plurality) same cycles)	1	<u>L5</u>
<u>L6</u>	L1 and (different temperature or different Tm)	1	<u>L6</u>
<u>L7</u>	multiplex near (PCR or polymerase chain reaction)	1296	<u>L7</u>
<u>L8</u>	L7 and (PCR cycles)	229	<u>L8</u>
<u>L9</u>	L8 and (different same (reaction near\$3 condition))	0	<u>L9</u>
<u>L10</u>	(PCR or polymerase chain reaction or RT-PCR or reverse transcriptase Polymerase chain reaction or polymerase chain amplification)	89525	<u>L10</u>
<u>L11</u>	L10 AND ((MULTIPLE OR PLURALS OR DIFFERENT OR MORE) NEAR CYCLE\$1)	2592	<u>L11</u>
<u>L12</u>	L10 AND (SEQUENTIAL NEAR CYCLE\$1)	80	<u>L12</u>
<u>L13</u>	L10 AND ((DIFFERENT OR DISTINCT OR SEPARATE) NEAR (TEMPERATURE OR TM))	2197	<u>L13</u>
<u>L14</u>	((L11 OR L12 OR L13) AND ((FIRST NEAR\$3 AMPLIFICATION)OR (FIRST NEAR\$3 PRIMER SET) OR (FIRST NEAR\$3 AMPLICON)))	0	<u>L14</u>
<u>L15</u>	((L11 OR L12 OR L13) AND (FIRST NEAR\$3 AMPLIFICATION))	0	<u>L15</u>
<u>L16</u>	((L11 OR L12 OR L13) AND (FIRST NEAR AMPLIFICATION))	250	<u>L16</u>
<u>L17</u>	((L11 OR L12 OR L13) AND (FIRST NEAR PRIEMR SET))	0	<u>L17</u>
<u>L18</u>	((L11 OR L12 OR L13) AND (FIRST NEAR PRIMER SET))	48	<u>L18</u>
<u>L19</u>	((L11 OR L12 OR L13) AND (FIRST NEAR AMPLICON))	28	<u>L19</u>
<u>L20</u>	L19 and (second near amplicon)	11	<u>L20</u>
<u>L21</u>	L18 and (second near primer set)	40	<u>L21</u>
<u>L22</u>	(L16 and (second near amplification))	125	<u>L22</u>
<i>DB=USPT,DWPI; PLUR=YES; OP=ADJ</i>			
<u>L23</u>	(L18 or L19 or L20 or L21 or L22) and (multiplex or multiplex amplification)	18	<u>L23</u>
<i>DB=USPT,USOC,EPAB,DWPI; PLUR=YES; OP=ADJ</i>			
<u>L24</u>	GUS near\$3 amplicon or 18S-rRNA near\$3 amplicon or tyrosinase near\$3 amplicon or CEA near\$3 amplicon	0	<u>L24</u>
<u>L25</u>	reverse transcriptase polymerase chain reaction or RT-PCR	7480	<u>L25</u>
<u>L26</u>	L25 same PCR primer set	14	<u>L26</u>
<u>L27</u>	L26 same polymerase	3	<u>L27</u>
<u>L28</u>	L27 and (single tube or single vessel or single cartridge)	0	<u>L28</u>
<u>L29</u>	L27 and ((single tube) or (single vessel) or (single cartridge))	0	<u>L29</u>
<u>L30</u>	L25 and ((single tube) or (single vessel) or (single cartridge))	168	<u>L30</u>
<u>L31</u>	L25 same((single tube) or (single vessel) or (single cartridge))	50	<u>L31</u>
<u>L32</u>	((single tube) or (single vessel) or (single cartridge)) same L2	13	<u>L32</u>
<u>L33</u>	L32 and L25	3	<u>L33</u>
<u>L34</u>	L25 and cartridge	536	<u>L34</u>
<u>L35</u>	L34 and compartments	98	<u>L35</u>

PubMed ID: 8158116

TITLE: Simultaneous amplification and detection of specific hepatitis B virus and hepatitis C virus genomic sequences in serum samples.

AUTHOR: Nedjar S; Mitchell F; Biswas R

CORPORATE SOURCE: Laboratory of Hepatitis, Center for Biologics Evaluation and Research, Food and Drug Administration, Bethesda, Maryland.

SOURCE: Journal of medical virology, (1994 Feb) 42 (2) 212-6.

Journal code: 7705876. ISSN: 0146-6615.

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DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals

ENTRY MONTH: 199405

ENTRY DATE: Entered STN: 19940526

Last Updated on STN: 19940526

Entered Medline: 19940517

AB A sensitive and specific **two-stage** polymerase chain reaction (PCR) technique was developed for the simultaneous **amplification** and detection of specific genomic sequences of hepatitis B virus (HBV) and hepatitis C virus (HCV) in serum samples. Initially, HCV-RNA was reverse transcribed to cDNA. This cDNA and DNA from HBV were then co-amplified using **primer pairs** derived from conserved regions of HBV and HCV nucleotide sequences. The specificity of PCR products was confirmed by liquid hybridization analysis using 32P end-labeled oligomer probes specific for the target HBV and HCV nucleotide sequences. Independent human serum samples, positive and negative by PCR for both HBV-DNA and HCV-RNA, were used as controls. We tested sera from nine donors, of which seven were reactive for HBsAg, anti-HBc, and anti-HCV (multiantigen test), one of whom was reactive for anti-HCV and anti-HBc, and one of whom was reactive for HBsAg and anti-HBc. The assay detected HBV- and HCV-specific genomic sequences in eight of eight sera reactive for both HBV and HCV serological markers and also in the serum that was reactive for HBV markers only.

Genotyping of hepatitis C virus by the polymerase chain
reaction: A comparison of two PCR-based methods using core
or NS5 primers.

AUTHOR: Hashimoto M.; Chayama K.; Tubota A.; Koida I.; Saitoh S.;
Arase Y.; Ikeda K.; Kobayashi M.; Kanda M.; Iwasaki S.;
Nakano A.; Takagi K.; Koike H.; Kumada H.

CORPORATE SOURCE: Liver Research Laboratoire, Department Gastroenterology,
Toranomon Hospital, Toranomon, Minato-Ku, Tokyo 105, Japan

SOURCE: International Hepatology Communications, (1995) 4/2
(67-71).

ISSN: 0928-4346 CODEN: IHCOEP

COUNTRY: Ireland

DOCUMENT TYPE: Journal; Article

FILE SEGMENT: 004 Microbiology
006 Internal Medicine
029 Clinical Biochemistry
048 Gastroenterology

LANGUAGE: English

SUMMARY LANGUAGE: English

AB The presence of hepatitis C virus subtypes and their clinical importance,
especially in the context of interferon sensitivity, are reported. In this
study, we compared two kinds of polymerase chain reaction (PCR
) -based methods - one step quick subtyping and **two stage**
subtyping procedure for the determination of genotypes. Using the mixed
primer sets derived from the core or NS5 region, we
amplified cDNA samples obtained from patients with a hepatitis C virus
infection. Of 116 samples tested, 91 (78%) showed identical genosubtypes
between these two methods and 19 (16%) showed different ones. Although
such discrepant results are considered to be related to the
characteristics of these two methods, further studies remain to
elucidated.

Sensitivity of **two-stage PCR**

amplification for detection of mycobacterium tuberculosis in paraffin-embedded tissues.

AUTHOR: Durmaz R.; Aydin A.; Durmaz B.; Aydin N.E.; Akbasak B.S.; Gunal S.
CORPORATE SOURCE: R. Durmaz, Department of Clinical Microbiology, Faculty of Medicine, Inonu University, Malatya, Turkey
SOURCE: Journal of Microbiological Methods, (1997) 29/2 (69-75).
Refs: 29
ISSN: 0167-7012 CODEN: JMIMDQ
PUBLISHER IDENT.: S 0167-7012(97)00022-5
COUNTRY: Netherlands
DOCUMENT TYPE: Journal; Article
FILE SEGMENT: 004 Microbiology
005 General Pathology and Pathological Anatomy
LANGUAGE: English
SUMMARY LANGUAGE: English

AB In order to improve the sensitivity of polymerase chain reaction (PCR) for the detection of mycobacterial DNA in paraffin-embedded tissues, a new approach with two sets of specific primers in **two-stage PCR** was employed in specimens obtained from tuberculosis patients and controls. Thirty-nine paraffin blocks selected from patients who had been diagnosed as having tuberculosis by radiological evaluations, histopathological findings, and clinical symptoms and signs including response to antituberculous treatment were examined. The control group consisted of 10 specimens from individuals that were proved to be negative for tuberculosis. After deparaffinization, lysis, phenol-chloroform extraction, and ethanol precipitation, the isolated DNA was amplified by PCR. Initially, all specimens were examined by the one-stage PCR using specific primers for 123-base pair (bp) fragment in IS6110 of mycobacterial DNA which yielded positive results only in 3 out of 39 (7.7%). In the **two-stage PCR** technique, 245 bp fragment of mycobacterial DNA was **amplified** at the first-step, then the PCR products were reamplified using the second specific **primer pairs** for 123-bp fragment. The true positivity of the **two-stage PCR** was 84.6% (33/39). The results indicate that **two-stage PCR** is more sensitive than one-stage (84.6% vs. 7.7%). All control specimens were negative by both PCR amplification methods, indicating that specificity of both methods was high. When the **two-stage amplification** was used, PCR positivity in the specimens obtained from different tissues was as follows: peritoneal and omental biopsies, 4/4; bone biopsies, 3/3; lymph node biopsies, 12/14; genito-urinary biopsies, 7/9; skin biopsies, 4/6; and one from each lung, breast, and pleural biopsies. PCR showed a good correlation with the granulomatous tissue reaction resulting in a 83.8% (31/37) positivity. The results indicate that the **two stage PCR amplification** can be used for detection of M. tuberculosis in paraffin-embedded tissues and is a useful technique in confirming tuberculosis in patients with clinically suspected disease who have acid-fast stain-negative.

(FILE 'HOME' ENTERED AT 15:27:41 ON 01 FEB 2005)

FILE 'MEDLINE, BIOTECHDS, EMBASE, BIOSIS, SCISEARCH, CANCERLIT, CAPLUS'
ENTERED AT 15:30:26 ON 01 FEB 2005

L1 202 S (PRIMER PAIR# OR PRIMER SET#) AND (TM OR EFFECTIVE TM)
L2 0 S L1 AND (TWO-STAGE (5A) (AMPLIFICATION OR PCR))
L3 187 S TWO-STAGE PCR OR TWO-STAGE AMPLIFICATION
L4 0 S L3 AND L1
L5 165 S L1 AND PCR
L6 14 S L5 AND MULTIPLEX PCR
L7 0 S L1 AND ((TWO-STEP OR TW- STAGE) (5A) PCR)
L8 4 S ((PRIMER PAIR# OR PRIMER SET#) (5A) (MELTING TEMPERATURE OR
L9 48 S ((PRIMER PAIR# OR PRIMER SET#) AND (MELTING TEMPERATURE OR E
L10 44 S L9 AND (PCR OR MULTIPLEX PCR OR POLYMERASE CHAIN REACTION OR
L11 21 DUP REM L10 (23 DUPLICATES REMOVED)

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